



Date: 17.12.2015

tail bleeding

LTK-TRT-19-A-EN
Version: A

This SOP replaces: Date: None
 Version: None

Reason for Change: None

Related SOPs:

Indication of Use: Analysis of cellular and non-cellular composition of venous blood

Aim of SOP: This protocoll describes how small volume blood samples are collected from the tail vein

- Distribution:**
1. Original: Server
 2. Copy: Animal facilities
 3. All Modul 1 certified Scientists

Attachments:


Generated
at: 31.08.2015

Checked and approved
at: 20.10.2015

by: Johannes vom Berg

by: Thorsten Buch

Responsible Persons: Researcher with Modul 1 after registration on animal license

 <p>University of Zurich UZH Institute of Laboratory Animal Sciences</p>	<p>Standard Operating Procedure</p> <p>SOP</p>	<p>Page 2 of 4</p>
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Method: bleeding

Min/Max amount:

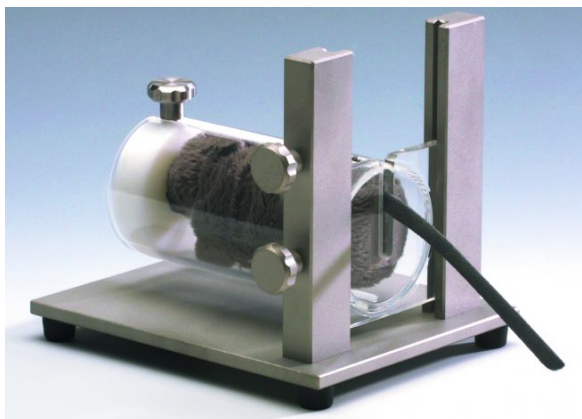
The maximum volume of blood collected is 1% of body weight (BW, 200 µl / 20 g mouse)
With 1% of BW bled, next bleeding after 2 weeks, with less than 0.25 % (50 µl / 20 g mouse)
after 48h. For immunological assays such as ELISA and flow cytometry 50 µl is sufficient

Storage of Material:

Heparinized whole blood collection tubes (orange cap, BD) and serum collection tubes (e.g. yellow cap, BD vacutainer SST tubes) are found in the injection lab B205 (as labelled on cupboards) and in the U211 (overhead cupboards)

Material:

1. scalpel, straight edge razor
1. heparinized whole blood collection tubes (orange cap, BD) or serum separator tubes (yellow cap, BD)
2. restrainer (1)



3. water in beaker at 45°C

Safety:

1. General rules for working with sharp tools (scalpels, syringes, scissors) have to be followed.
2. Follow the rules of the animal house



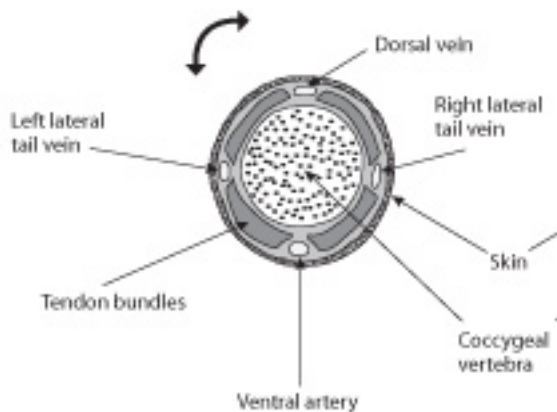
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Method Description:

2. Warm the tail of the mouse in 45°C warm water.
3. Restrain the mouse in the physical restrainer device or anaesthetize the mouse.
4. Using a scalpel, straight edge razor, or sharp scissors, quickly nick the lateral tail vein (see transverse section below, 2) at the distal part of the tail close to the tip.



5. Collect blood in a capillary tube as drops appear. Do not attempt to increase blood flow by rubbing the tail from the base to the tip, as this will result in leukocytosis (increased white blood cell count)
6. Collect up to 1% of body weight (200 µl / 20 g mouse) of blood into a collection tube (see materials)
7. take a sterile swab and apply pressure on the wound to stop the bleeding
8. Gently release mouse from the restrainer and place it back into the home cage

Documentation:


Lab Journal, iRATS - put animal into experiment and project, severity 1

Problem management:

In case of serious adverse events, contact supervisor, lab head or vet.

Sample storage:

- Do not freeze samples before you have separated serum. Freeze serum at -80°C for later analysis by ELISA or WB
- Keep blood samples for FACS at 4°C until analysis.

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Literatur:

1. http://www.mouse-restrainer.de/index_en.html
2. The Laboratory Mouse, Chapter 32: routes of administration, Shinya Shimizu, figure 32.12 a, Copyright 2004, Elsevier ISBN 0-1233-6425-6
3. Dürschlag M, Würbel H, Stauffacher M, Von Holst D. Repeated blood collection in the laboratory mouse by tail incision--modification of an old technique. *Physiol Behav.* 1996 Dec;60(6):1565-8. PubMed PMID: 8946506.
4. Diehl KH, Hull R, Morton D, Pfister R, Rabemampianina Y, Smith D, Vidal JM, van de Vorstenbosch C; European Federation of Pharmaceutical Industries Association and European Centre for the Validation of Alternative Methods. A good practice guide to the administration of substances and removal of blood, including routes and volumes. *J Appl Toxicol.* 2001 Jan-Feb;21(1):15-23.