

 <b>University of Zurich</b> Institute of Laboratory Animal Sciences	<b>Standard Operating Procedure</b>  <b>SOP</b>	Page 1 of 3
<b>Date: 08.11.2022</b>	<b>Adoptive transfer tumor</b>	<b>LTK-RES-49-B-EN</b> <b>Version: B</b>

<b>This SOP replaces:</b> Date: A	
<b>Reason for Change:</b> Response to veterinary office	
<b>Related SOPs:</b> SOP LTK-TRT-7 iv injection SOP LTK-TRT-14 CO <sub>2</sub> euthanasia	
<b>Indication of Use:</b> Determination of tumor responses triggered by adoptively transferred cells	
<b>Aim of SOP:</b> This protocol describes how to perform mouse treatment by adoptive transfer of cells	
<b>Distribution:</b> 1. Server 2. Animal facility 3. Group vom Berg	
<b>Attachments:</b>	
Generated at: 07.11.2022	Checked and approved at: 08.11.2022
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**Date: 08.11.2022**

**Adoptive transfer tumor**

**LTK-RES-49-B-EN  
Version: B**

**Responsible Persons:**

- 1) The researcher mentioned on the respective scoring sheet.
- 2) Any person with Module 1 and registered on animal permit.

**Method:** Preparation of cells for adoptive transfer

**Principle of Method:** Cells for adoptive transfer are purified from organs of donor mice and prepared for injection into tumor-bearing animals.

**Material to be used:**

PBS  
50 mL sterile polypropylene tubes  
Pipettes - 5 ml, 10 ml, 25 ml  
Syringe 10 ml  
70 µm cell strainer  
Magnetic cell separation kit

**Storage of Material:**

Follow the individual data sheets for all chemicals used.

**Machine:**

Centrifuge  
Magnets for magnetic cell separation

**Safety:**

1. General rules for working with sharp tools (scalpels, syringes, scissors) have to be followed.
2. Follow the rules of the animal house.
3. Follow the rules of working with toxic chemicals.



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**Method Description:**

Perform all cell preparation steps aseptically in a biosafety cabinet. Keep cells cold (4°C). Use cold PBS and keep cells on ice when practical.

1. Euthanize donor mouse and collect the spleen (see **SOP LTK-TRT-14 CO2 euthanasia**)
2. Place spleen in a strainer inside a petri dish. Squash spleen in the Petri dish by pressing several times with the hard end of a 10 ml syringe plunger. Place a fresh 70 µm cell strainer in a 50 ml tube. Collect all PBS and squashed tissue from the Petri dish into the cell strainer.
3. Spin down cells in 50 ml tubes for 5 minutes at 350 g. Discard supernatants.
4. Purify cells using magnetic cell separation. E.g., for purification of OT-I cells use CD8<sup>+</sup> T cell purification kit, for purification of OT-II cells use CD4<sup>+</sup> T cell purification kit.
5. Count the cells and resuspend in PBS to reach up 10<sup>7</sup> cells per mouse and volume up to 5 µl/g of mouse body weight.
6. Inject cells i.v. according to **SOP LTK-TRT-7 iv injection**

**Documentation:**

Server, appropriate project folder.

**Problem management:**

Report any adverse event to your supervisor